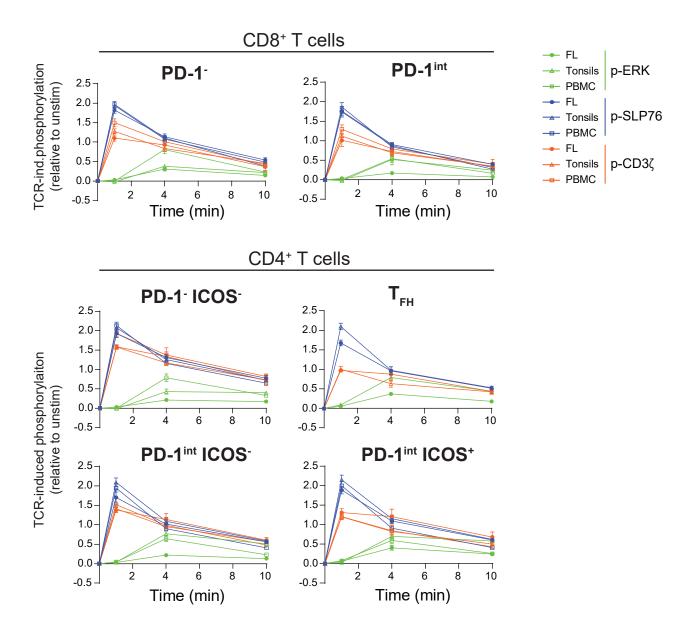
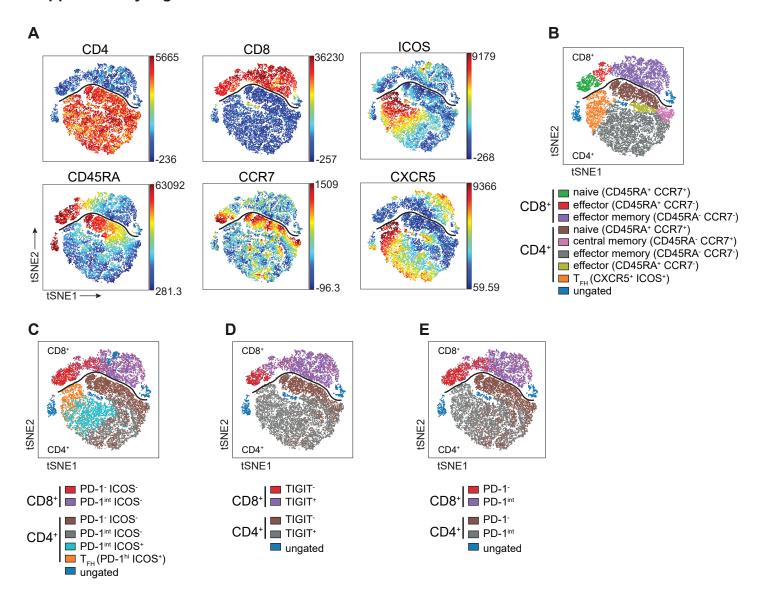


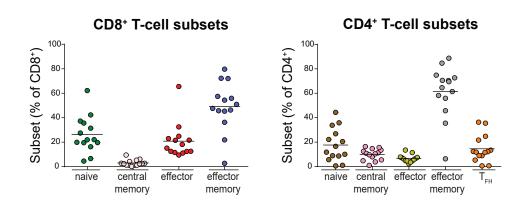
Supplementary Figure 1. IL-21 production is reduced in CD4 FL T cells. Single cell suspensions from FL LN and healthy donors (tonsils and PBMC) were cultured with or without PMA and ionomycin for a total of 6 hours, with GolgiPlug present for the last 4 hours. Levels of intracellular IL-4 and IL-21 were measured by fluorescence flow cytometry. Each data point represents a single donor. Statistical differences were calculated using Mann-Whitney non-parametric test; *p < 0.05, **p < 0.01, ***p < 0.001.



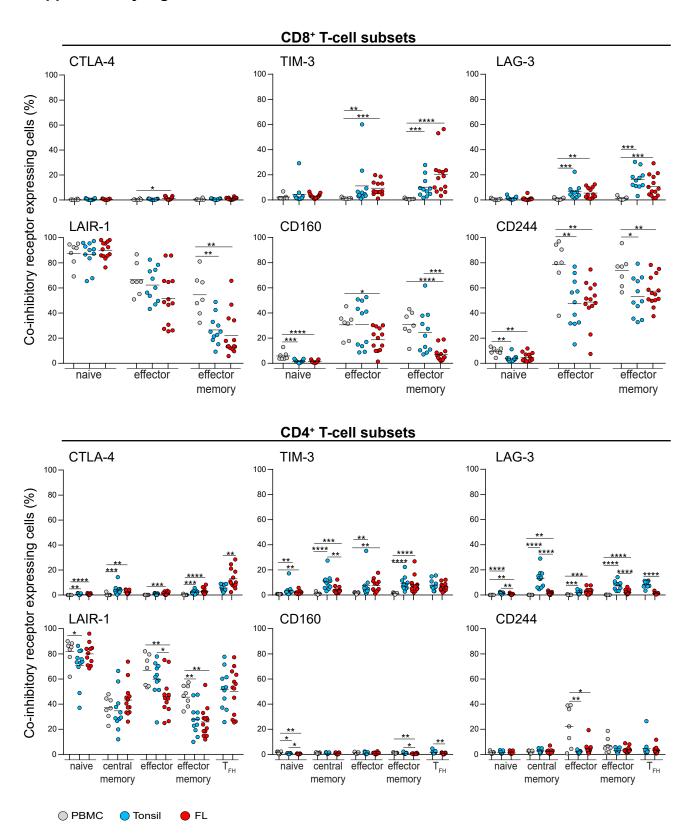
Supplementary Figure 2. TCR-induced signaling effectors have different kinetic. Single cell suspensions from FL LN (n=9), tonsils (n=11) and PBMC (n=9) were cultured with or without α -CD3+ α -CD28 antibodies for 2 minutes, followed by avidin crosslinking for 1, 4 or 10 minutes and then assayed for TCR-induced phosphorylation of CD3 ζ , SLP76 and ERK using phospho-flow cytometry. Diagrams show kinetic of the three signaling effectors in CD8 and CD4 T-cell subsets as median fold change (FC) induction relative to unstimulated cells. Mean FC \pm SEM.



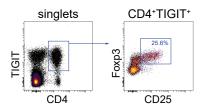
Supplementary Figure 3. Identification of T-cell subsets. Single cell suspensions from FL were analyzed by fluorescence flow cytometry. A viSNE analysis was performed on the CD3⁺ cells, based on expression of CD4, CD8, CXCR5, ICOS, CD45RA and CCR7. Figure shows viSNE map of 4 FL samples combined. The manually added line in the viSNE plots marks the distinction between CD8 and CD4 T cells. Color represents fluorescence intensity with separate scale for each marker, with max set to highest measured signal. (A) Intensity of markers included in viSNE analysis and used for gating of conventional T-cell subsets (corresponding to subsets used in Figure 4). (B) Each gated T-cell subset, identified in (A), is shown with a separate color. (C) T-cell subsets were gated on biaxial plots based on expression of PD-1 and ICOS as in Figure 1, and shown on the viSNE map. (D) T cells were gated on biaxial plots based on expression of CD4/CD8 and TIGIT and shown on viSNE map. (E) T cells were gated on biaxial plots based on expression of CD4/CD8 and PD-1 and shown on viSNE map.



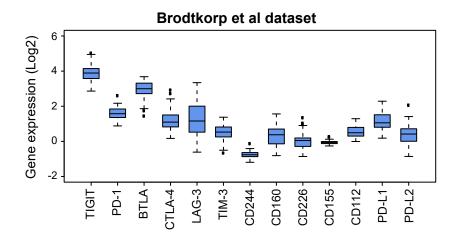
Supplementary Figure 4. Distribution of T-cell subsets in FL LN. Single cell suspensions from FL LN (n = 14) were analyzed by fluorescence flow cytometry. Conventional subsets of naive T cells, T central memory cells, T effector cells, T effector memory cells and T follicular helper cells ($T_{\rm FH}$) were gated on biaxial plots, based on differential expression of CD4, CD8, CXCR5, ICOS, CD45RA and CCR7. Each data point represents a single donor.

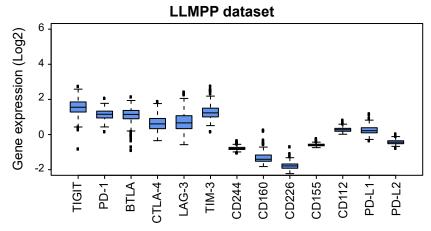


Supplementary Figure 5. Expression patterns of co-inhibitory receptors in CD8 and CD4 T-cell subsets. Co-Inhibitory receptor expression was analyzed by 11-parameter flow cytometry in single cell suspensions from FL LN and non-tumoral controls (tonsils and PBMC). Each data point represents a single donor. Statistical differences were calculated using Mann-Whitney non-parametric test *p < 0.05, **p < 0.01, ***p < 0.001, ****p < 0.0001.

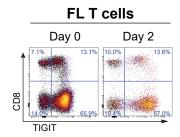


Supplementary Figure 6. Contribution of Tregs among CD4***TIGIT*** **T cells** Single cell suspensions were analyzed by flow cytometry. Plots show one representative FL sample.



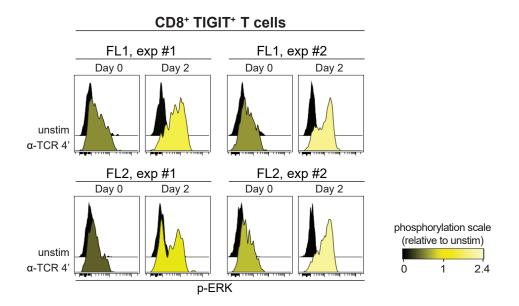


Supplementary Figure 7. TIGIT is a highly expressed co-inhibitory receptor in FL. Boxplots represent gene expression in 44 pre-treatment FL biopsies (Brodtkorb et al.) (upper panel), and 194 pre-treatment FL biopsies (Dave et al.) (lower panel). Values are median-centered across patients.



Supplementary Figure 8. TIGIT expression is stable over time.Single cell suspensions from FL LN were analyzed by flow cytometry.

TIGIT expression was measured at day 0 and after 48 h in vitro culture.



Supplementary Figure 9. Recovery of TCR-induced signaling in TIGIT⁺CD8⁺ FL T cells is robust. Single cell suspensions from FL LN were assayed for TCR-induced signaling and analyzed by phospho-flow cytometry at day 0 and after 48 hour *in vitro* culture. Signaling was induced using α -CD3+ α -CD28 antibodies for 2 minutes, followed by avidin crosslinking for 4 minutes. The same FL specimens (n = 2) were analyzed in two separate experiments. The histograms show median fold change (FC) induction relative to unstimulated cells, using arcsinh transformed data.