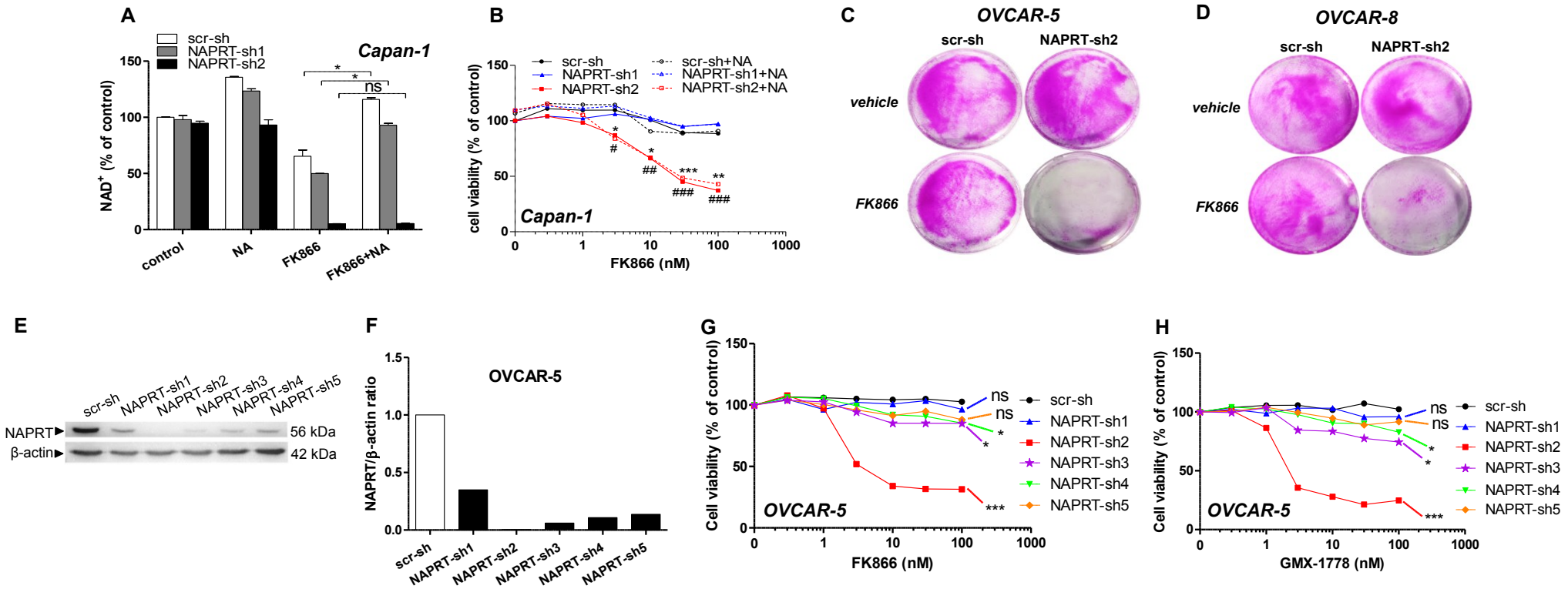

Figure S3. **NAPRT silencing sensitizes pancreatic and ovarian cancer cells to FK866**. A, Capan-1 cells were transduced with the NAPRT-sh1, -sh2 or with a scrambled shRNA. 10^5 Capan-1 cells expressing the scr-shRNA, NAPRT-sh1 or NAPRT-sh2/well were plated in 6-well plates, allowed to adhere overnight and then treated for 24 h w/ or w/o 100 nM FK866 in the presence or absence of 10 μM NA. Thereafter, cells were used for NAD^+ determination. B, 2 x 10^3 Capan-1 cells expressing the scr-shRNA, NAPRT-sh1 or NAPRT-sh2/well were plated in 96-well plates, allowed to adhere overnight and then incubated for 72 h w/ or w/o FK866 at the indicated concentrations w/ or w/o 10 μM NA. Thereafter, cell viability was detected with SRB. C, D, 4 x 10^4 OVCAR-5 and OVCAR-8 cells were plated in 60 mm Petri dishes. 24 h later, 100 nM FK866 or vehicle DMSO were added. 72 h later, the culture plates were fixed and the plates were stained with SRB, dried and photographed. E, F, OVCAR-5 cells were transduced with the NAPRT-sh1-5 and used for protein lysate generation. NAPRT and β-actin were detected by immunoblotting. Band intensities were quantified and the NAPRT/actin ratio was calculated (F). G, H, 2 x 10^3 OVCAR-5 cells expressing the scr-shRNA or one of the NAPRT-sh1-5/well were plated in 96-well plates, allowed to adhere overnight and then incubated for 72 h w/ or w/o FK866 or GMX-1778 at the indicated concentrations. Thereafter, cell viability was detected with SRB. A, Data are presented as means ± SD of three separate experiments. B, G, H, Data are presented as means of three separate experiments. C-F, One representative experiment out of three is presented. ns: non-significant; *: p<0.05; **: p<0.01; ***: p<0.001; #: p<0,05; ##: p<0,01; ###: p<0,001. In G, H, the statistical analysis refers to the cell viability observed in response to 100 nM FK866 or GMX-1778. In B, hashtag symbols refer to statistical significance of NAPRT-sh2+NA samples.