**Supplemental Materials and Methods on Staining Protocols for Westerns and Flow Cytometry**

Western blot analysis: Cells (SVEC, H460, A549) were lysed in 1% Triton X-100 lysis buffer. The protein concentration of cleared supernatants was determined using BCA reagent (Thermo Fisher Scientific) and SoftMax Pro software (SpectraMax M3, Molecular Devices). Equivalent amounts of protein were loaded for each lysate on a 4-12% Bis-Tris NuPAGE gel (Life Technologies). Proteins were transferred to nitrocellulose membranes, blocked for 1h in 5% milk (in 1X Tris Buffered Saline with Tween; TBST), then probed overnight in 5% milk with anti-EGFR antibody at 4°C with rocking (Cell Signaling Technologies) and with anti-β-actin antibody (internal loading control) for 30min-1h at room temperature. Following primary antibody incubation, membranes were washed with 1X TBST for at least 1h with multiple changes of solution. Membranes were then probed in 5% milk with goat anti-rabbit HRP (to visualize EGFR) and goat anti-mouse HRP (to visualize β-actin) conjugated secondary antibodies (BioRad) for 30min-1h. Following a final wash in 1X TBST for at least 1h with multiple changes of solution, membranes were placed in enhanced chemiluminescence reagent (GE Healthcare) for 1min, excess reagent was removed, and membranes were exposed to X-ray film. Film was scanned and blot image was composed in Adobe Photoshop CS5.

Flow cytometry: Treated or control H460-RFP tumors (2-5 pooled/condition) were excised, then minced and rocked (15 min, 37°C) in a solution of collagenase/dispase/HBSS. DNase was added and dissociation continued for an additional 15 min. Cells were centrifuged (1000 rpm) and resuspended in 1X Red Blood Cell Lysis Buffer (eBioscience). After stopping this reaction with an excess of ice-cold HBSS, cells were centrifuged, resuspended in staining buffer (3% FBS in HBSS), filtered (70µm nylon mesh; BD Biosciences), counted and resuspended to a final concentration of 2x107 cells/ml in staining buffer. Staining was performed with Annexin V (labeling apoptotic cells; eBioscience) and DAPI (labeling necrotic cells; Life Technologies). Samples were analyzed on a LSRII Flow Cytometer using Diva software (Becton Dickinson, Mountain View, CA). Data were analyzed in FlowJo (Treestar, San Carlos, CA).