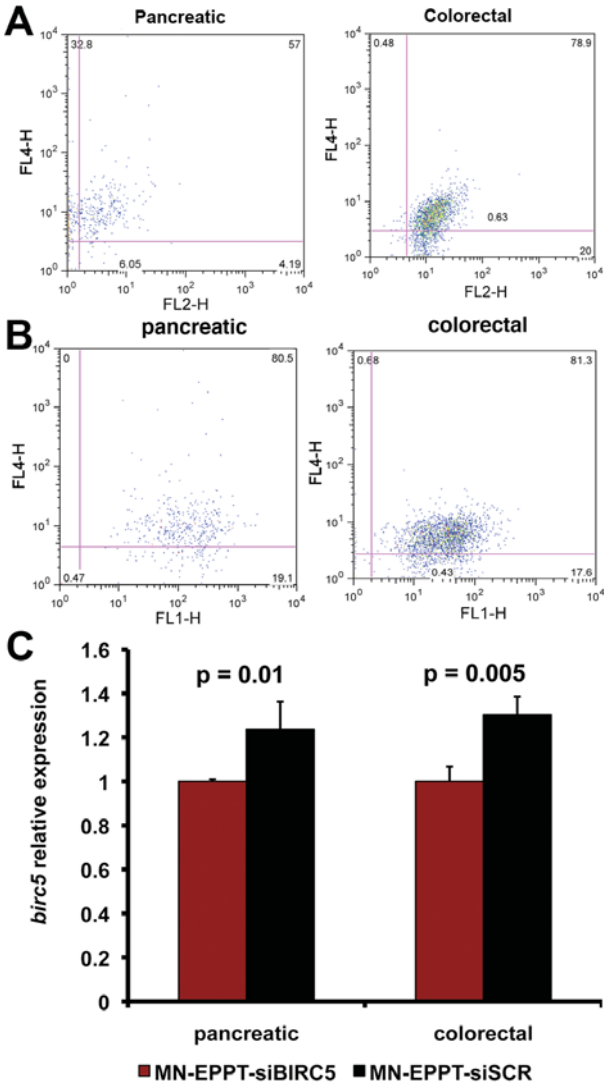
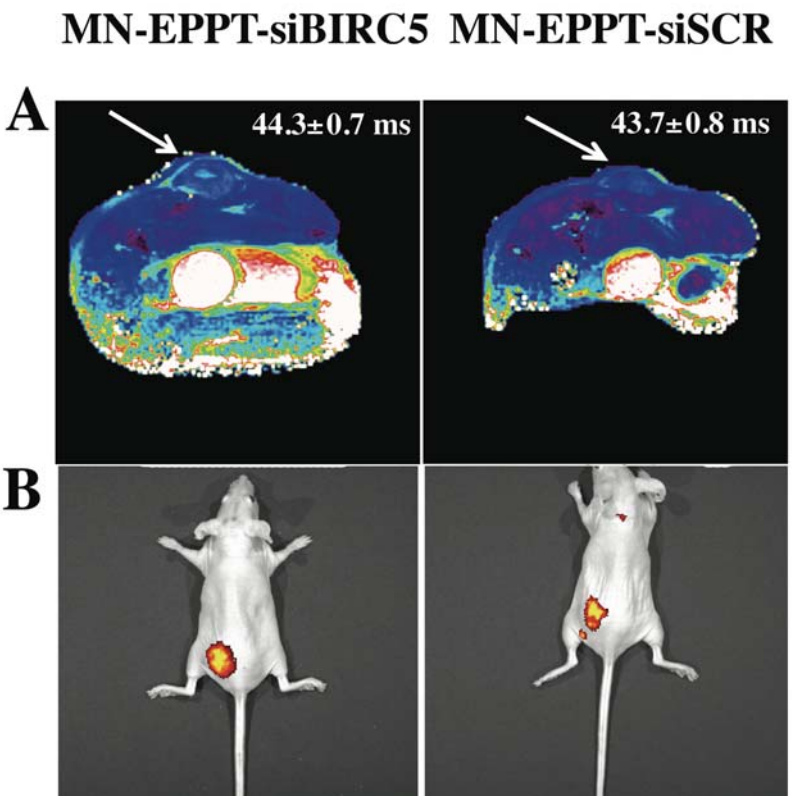


SUPPLEMENTAL FIGURE 1. In vitro testing of MN-EPPT-siRNA uptake and silencing efficacy in human pancreatic (CAPAN-2) and colorectal (LS-174T) adenocarcinoma cells. **(A)** Flow cytometry to assess nanodrug uptake. Representative FL2 (Dy547, siRNA) vs. FL4 (Cy5.5, MN) dot plots showing that the cells were labeled with the nanodrug. The cellular co-localization between fluorescence in the two channels indicated stability of the nanodrug. **(B)** Flow cytometry to assess nanodrug uptake as a function of uMUC-1 positivity. The cellular co-localization between fluorescence in the

FL4 (Cy5.5, MN) and FL1 (FITC, uMUC-1-specific antibody) channels suggested that the nanodrug uptake by the cells is representative of uMUC-1 abundance. **(C)** qRT-PCR of human pancreatic (CAPAN-2) and colorectal (LS-174T) adenocarcinoma cells incubated with MN-EPPT-siBIRC5 or control probes. There was a significant knock-down of *birc5* mediated by MN-EPPT-siBIRC5 relative to the MN-EPPT-siSCR control (n = 4).



SUPPLEMENTAL FIGURE 2. In vivo tumor uptake of MN-EPPT-siBIRC5 vs. MN-EPPT-siSCR. **(A)** T2 weighted MR imaging. There was no difference between the levels of tumor-associated signal loss mediated by the two probes. The T2 relaxation times of the two groups were not significantly different (44.3±0.7 vs 43.7±0.8 ms; n = 4). **(B)** In vivo near-infrared optical imaging. There was no visible difference between the levels of tumor-associated fluorescence mediated by the two probes.